

# Effects of temperature on embryonic and larval development and growth in the natterjack toad (*Bufo calamita*) in a semi-arid zone

D. Sanuy, N. Oromí & A. Galofré

Sanuy, D., Oromí, N. & Galofré, A., 2008. Effects of temperature on embryonic and larval development and growth in the natterjack toad (*Bufo calamita*) in a semi-arid zone. *Animal Biodiversity and Conservation*, 31.1: 41–46.

## Abstract

*Effects of temperature on embryonic and larval development and growth in the natterjack toad (Bufo calamita) in a semi-arid zone.*— Temperature affects the duration of embryonic and larval periods in amphibians. Plasticity in time to metamorphosis is especially important in amphibian populations of Mediterranean semi-arid zones where temperatures are high and precipitation is low, increasing the rate of pond desiccation. In order to test the influence of water temperature on the larval development and growth of the natterjack toad (*Bufo calamita*), we collected two spawns in a semi-arid zone at Balaguer (Lleida, NE Iberian peninsula). Approximately 50 ( $\pm 10$ ) eggs (stage 14–16) were raised in the lab at different temperature conditions: 10, 15, 20, 22.5 and 25°C with 12:12 photoperiod. The results show a lengthening of development time with decreasing temperatures and a better survival performance of *B. calamita* to high temperatures. However, mean size at metamorphosis was not different across treatments, thus, suggesting that this population of *B. calamita* requires a minimum size to complete the metamorphosis. This study is the first approach to examine the effects that climatic factors have on the growth and development of *B. calamita* in semi-arid zones.

Key words: Amphibians, Development, Growth, Semi-arid zone, Temperature.

## Resumen

*Efectos de la temperatura sobre el desarrollo y crecimiento de embriones y larvas en el sapo corredor (Bufo calamita) en zonas semiáridas.*— La temperatura afecta la duración de los periodos embrionarios y larvarios de los anfibios. La plasticidad en el momento de la metamorfosis es especialmente importante en las poblaciones de anfibios del Mediterráneo y de zonas semiáridas donde las temperaturas son elevadas y las precipitaciones son escasas, lo que incrementa la desecación de las charcas. Con el fin de comprobar la influencia de la temperatura del agua en el desarrollo de las larvas y crecimiento de la sapo corredor (*Bufo calamita*), se recolectaron huevos de dos puestas procedentes de una zona semiárida en Balaguer (Lleida, NE península ibérica). Aproximadamente unos 50 ( $\pm 10$ ), huevos (etapa 14–16) se sometieron en el laboratorio a diferentes condiciones de temperatura: 10, 15, 20, 22,5 y 25°C con un fotoperíodo 12:12. Los resultados muestran un alargamiento del tiempo de desarrollo con la disminución de las temperaturas y una mejor supervivencia de *B. calamita* a temperaturas mas altas. Sin embargo, el tamaño medio de los ejemplares metamórficos no fue diferente entre los tratamientos, lo que sugiere que esta población de *B. calamita* requiere un tamaño mínimo corporal para completar la metamorfosis. Este trabajo es un estudio preliminar para examinar los efectos que los factores climáticos tienen en el crecimiento y el desarrollo de *B. calamita* en las zonas semiáridas.

Palabras clave: Anfibios, Desarrollo, Crecimiento, Zonas semiáridas, Temperatura.

(Received: 5 VI 07; Conditional acceptance: 24 VII 07; Final acceptance: 24 IX 07)

Delfí Sanuy, Neus Oromí & Aina Galofré, Escola Tècnica Superior d'Enginyeria Agrària, Dept. de Producció Animal (Fauna Silvestre), Univ. of Lleida, Av. Rovira Roure 177, 25188 Lleida, Catalonia, Spain.

Corresponding author: D. Sanuy. E-mail: dsanuy@prodan.udl.es

## Introduction

Populations of ectotherm animals have a strong dependence on ambient temperature because they do not have an efficient mechanism for physiological thermoregulation (Brattstrom, 1963). Climatic variation is an important selective factor for life history trait differentiation, so populations of ectotherms are expected to diverge in their thermal optima for development and growth if they are exposed to different temperature environments. Amphibians have been widely used as model systems for the study of physiological ecology and temperature adaptation due to the easy manipulation of tadpoles in control experiments (Feber & Burggren, 1992).

In temporary ponds, time to metamorphosis is influenced by temperature and duration of the larval stage is highly plastic, especially in semi-arid environments where rainfall and pond duration are unpredictable (Newman 1989; Tejedo & Reques, 1994). In these environments, larvae need to accelerate metamorphosis when ponds dry out and delay the process when ponds dry later in the season. When pond duration is extremely short larvae should be the most adaptive strategy. It would be to express not a plastic but a quick canalized development (Tejedo & Reques, 1994). Predation and pond desiccation have been identified as major causes of larval mortality in permanent and transient aquatic environments, respectively (Brockelman, 1969; Calef, 1973; Smith, 1983; Newman, 1987).

In anuran populations, larger metamorphs may exhibit higher terrestrial survival since they can cope with different stressors such as predators and desiccation (Reques & Tejedo, 1997; Altwegg & Reyer, 2003). Smith-Gill & Berven (1979) considered that low temperatures retard differentiation more than growth and increase the stage-specific size. Therefore, larvae growing at cold temperatures have prolonged developmental periods but they may increase their size at metamorphosis. This phenomenon is considered a general rule for ectotherms (Atkinson, 1994, 1996). This trade-off in fitness, mediated by the influence of temperature, largely conditions the expression of an optimal phenotype at metamorphosis in amphibians and other aquatic organisms with complex life cycles (Etkin, 1964; Smith-Gill & Berven, 1979; Semlitsch et al., 1988; Hayes et al., 1993).

In this study, we examined the influence of temperature on the development and growth of the natterjack toad (*Bufo calamita*). The study area is a semi-arid zone with a marked deficit of precipitation and high temperatures. Previous studies of the species in the study zone suggest that *B. calamita* adults (Miaud et al., 2000) presents different terrestrial behaviour with respect to other natterjack populations in other areas of Europe. In this study we focus on the response of natterjack larvae to increasing temperatures and analyze their metamorphic response.

## Material and methods

The study area (Balaguer, 41° 46' N, 0° 46' E) is a semi-arid zone (Conesa et al., 1994) located in an agricultural setting of winter cereal fields. Mean annual temperature is about 14.5°C (January = 0.2°C and July = 32.2°C) and the mean annual rainfall is 400 mm (Balaguer meteorological station). The soil is poor and contains a high proportion of clay and lime. The evapo-transpiration is 805 mm (Thornthwaite & Mather, 1955). All these factors determine a harsh terrestrial environment with a particular flora and fauna that can tolerate such conditions (Conesa et al., 1994). As suitable breeding habitats for natterjack are temporary, tadpoles have to develop in a few days to complete the metamorphosis.

We examined the effects of temperature on size and time to metamorphosis in the natterjack toad, *Bufo calamita* (*Epidalea calamita* according to Frost et al., 2006), a common amphibian species in the study area. This species breeds in seasonal ponds. Breeding activity can occur throughout the year after rainfalls, except in the colder months, with an explosive peak in the periods of more intensive rains (Sanuy, unpublished data).

Two egg clutches were collected (approximately 100 eggs/spawn) in a temporal pond in the study area. At the lab where they were placed in a controlled temperature chamber (with an error of  $\pm 0.5^\circ\text{C}$ ) and immediately exposed to five different temperatures under a constant 12 hours light / 12 hours dark photoperiod. The temperature treatments were: 10, 15, 20, 22.5 and 25°C. At the start of the experiment eggs were near Gosner stage 16 of development (14–16; Gosner, 1960). Approximately 50 eggs ( $\pm 10$  eggs) from each spawn were raised in plastic recipients containing 1.3 L of dechlorinated water. Each spawn was replicated twice per experimental temperature camera. Larvae were fed fish food and the excess was removed daily. Water was changed every three days.

During the experiment the stage of development was checked and photographed every 12 hours. The total body length (BL) of each larva was estimated using the program coreIDRAW 11. The stage of development and BL of larvae in each sample were measured on days 1, 5 and 19 of the experiment. Stage-size was measured for each spawn at each temperature condition, at stages: 16, 25 and 46 (Gosner, 1960). Due to development time differences between individuals of the same spawn, the developmental stage was considered to have changed when 70% of larvae in the same sample had reached a particular stage. After the experiment was completed (experimental time: 150 days), the surviving larvae were returned to the pond where they were collected.

The relationship between the stage of development (dependent variable) and time at each temperature and for each spawn was analysed using two-way ANOVA (PROC GLM version 9 SAS). The same type of analysis was also used to measure stage-size and the relationship between size and

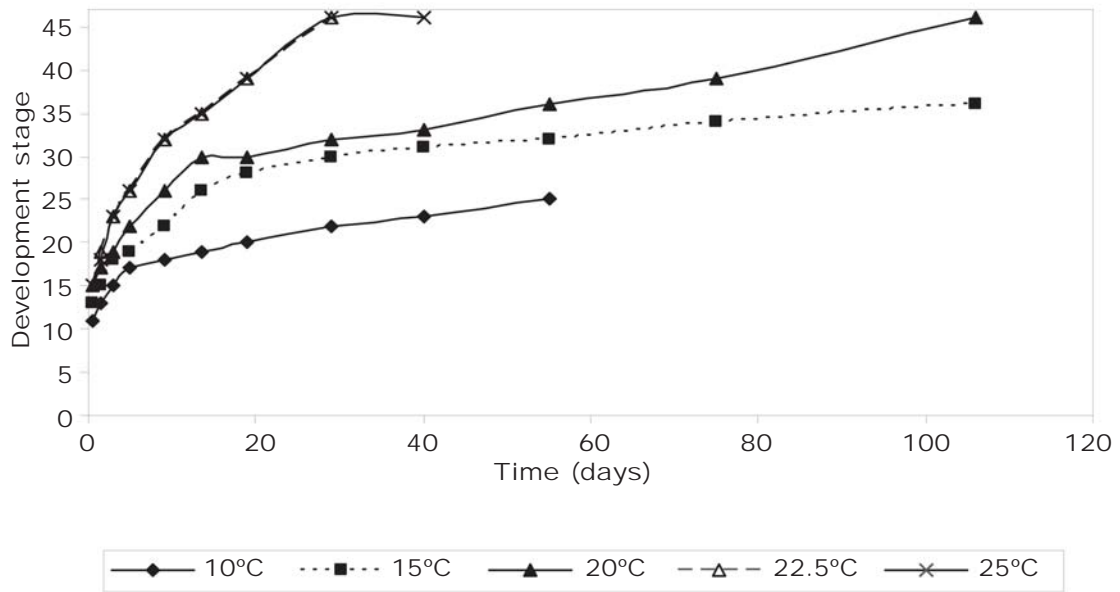


Fig. 1. Effect of temperature on *Bufo calamita* larval development (Gosner stages; Gosner, 1960).

Fig. 1. Efecto de la temperatura en el desarrollo larvario de *Bufo calamita* (estadios de Gosner; Gosner, 1960).

time. The significance level was set at  $\mu = 0.05$ . All data are expressed as mean  $\pm$  SD.

Results

Development time and survival.

Development time increased with decreasing temperature (fig. 1). Developmental stage differed for each temperature condition (5 day:  $F = 71.20$ ;  $p < 0.001$ , 19 day:  $F = 339.5$ ;  $p < 0.001$ ) and spawn (only in the 5 first days;  $F = 0.20$ ;  $p < 0.001$ ). Similarities in development were only observed between 22.5°C and 25°C (post hoc Tukey test, 5 day =  $p = 0.99$ , 19 day = 0.99), with Individuals in both temperatures metamorphosing in 23 days (stage 42). The coldest temperature treatment (10°C) had a negative effect on survival: tadpoles stopped development at stage 23 (55 days), dying in 10 days. In the 15°C treatment tadpoles stopped developing at stage 35 (90 days) and they survived without further development until the last day of the experiment. The larval period of tadpoles reared at 20°C was on average 88 days (stage 42).

Size at development stage

The size increased at each development stage. We found differences in the total size between temperatures at stage 25 (fig. 2, table 1). In stage

39 tadpoles attained the maximum body size and we found differences between 20, 22.5 and 20 and 25 temperature treatments but not between 22,5 and 25°C (table 2). However, all individuals completed the metamorphosis with a similar size ( $7.6 \pm 1.2$  mm;  $F = 0.08$ ;  $p = 0.92$ ) across temperature treatments.

Discussion

Temperature affects development and growth and is an important factor for climate selection (Berven & Gill, 1983). As expected, in our study, development time increased with decreasing temperature. Development time is very important for *Bufo calamita* in the study area because the breeding sites are temporary ponds with a high risk of desiccation due to high water temperatures and shallowness. Pond temperature plays a major role in determining the duration of the larval period in amphibians (Loman, 2002). Our results support other laboratory studies along these lines that have shown the effects of lowered water level on anuran development, presumably an adaptation to survival in drying ponds (Tejedo & Reques, 1994; Loman, 1999; Ryan & Winne, 2001). If pond drying increases temperature, our findings could be considered the result of drying effects. However, the analysis of effects of pond drying is complicated

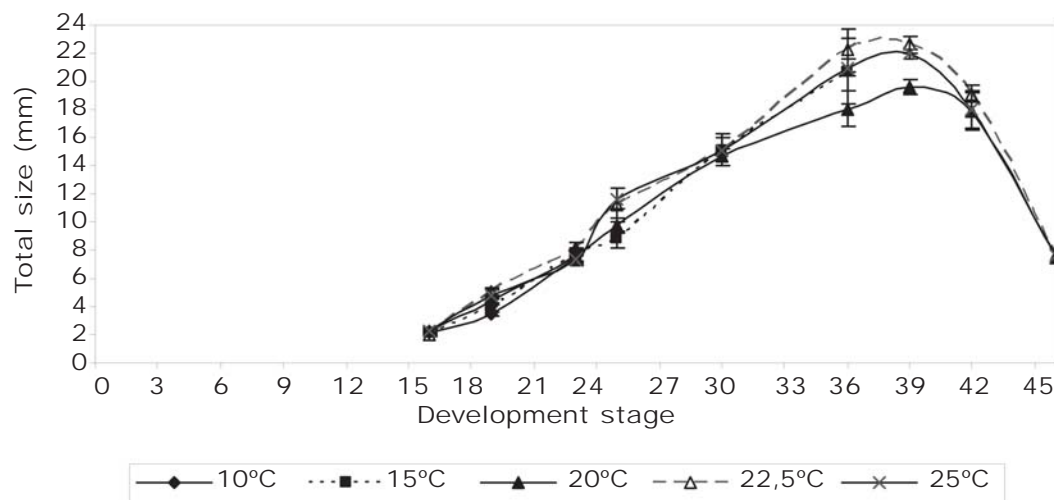


Fig. 2. Effect of temperature on larval size variation across developmental stages in *Bufo calamita*.

Fig. 2. Efecto de la temperatura sobre la variación del tamaño de las larvas, a través de todas las fases de desarrollo de *Bufo calamita*.

Table 1. Effect of temperature on the total size at stage 25 (Gosner, 1960). Two-way factorial ANOVA.

Tabla 1. Efecto de la temperatura sobre el tamaño total del estadio 25 (Gosner, 1960). ANOVA factorial bidireccional.

Source	df	F	P
Temperature	3	10.11	0.0017
Spawn	1	0.87	0.3710

because this factor interacts with others, such as pond temperature and tadpole density (Loman, 2002). Other authors, studying *B. calamita*, did not find evidence in the response to desiccation *per se*, suggesting that development may differ under physiological constraints in relation to habitat types (Brady & Griffiths, 2000).

Laboratory studies have shown a pattern in which tadpoles develop and grow more slowly but metamorphose with a longer body size at low temperatures (Etkin, 1964; Smith-Gill & Berven, 1979; Hayes et al., 1993). This result was not found in our study. Individuals raised at different temperatures attained metamorphosis at different times but contrarily to expected, size at metamorphosis was similar for all the treatments.

Table 2. Differences in maximum size reached due to temperature conditions at stage 39 (Gosner, 1960): PT. Post hoc Tukey test; T. Temperature; P. Probability.

Tabla 2. Diferencias en el tamaño máximo alcanzado en la fase 39, debido a las condiciones de la temperatura (Gosner, 1960): PT. Test post hoc de Tukey; T. Temperatura; P. Probabilidad.

	PT	T	P
T	LS mean	15 vs 20	0.3746
		15 vs 22.5	0.0074*
15	8.7792	15 vs 25	0.0024*
20	9.7645	20 vs 22.5	0.1201
22.5	11.2080	20 vs 25	0.0372*
25	11.6255	22.5 vs 25	0.8891

Therefore, higher temperature treatments determined both higher developmental rates and higher growth rates. All individuals that reached metamorphosis had the same body size. These results suggest that the population of *B. calamita* in the study zone requires a minimum size to complete metamorphosis.

Metabolic enzyme systems present high temperature sensitivity in an optimal range so growth and development can only proceed within a thermal window (e.g. Randall et al., 1997). The optimal temperature seems to be the higher temperatures (25 and 22.5°C) as tadpoles reached metamorphosis in a shorter time and with larger size, thus with potential higher fitness. The results of this study do not allow us to determine higher limits of temperature tolerance for the species. The air temperatures in the study area can reach maximums of about 40°C in summer, considerably higher than the maximum temperature used in our experiments. We cannot therefore determine a critical thermal maximum (CTM). On other hand, the minimum temperature that tadpoles needed to reach metamorphosis was 20°C, which is a higher minimum temperature compared to the temperature tolerance in other anuran species in Europe (for example: *Rana arvalis* Loman, 2002; *Rana temporaria* Laugen et al., 2003a, 2003b). Tadpoles attained a maximum size at stage 39 and decreased thereafter due to metamorphosis. The different sizes of larvae between temperatures at this stage, larger at higher temperatures, could be explained by the fact that the metabolism of growth factors is affected by temperature (Álvarez & Nicieza, 2002). Higher temperatures are well known to accelerating larval growth and development (Hayes et al., 1993). Growth rate depends on development rate, and both are functions of circulating hormone levels, which are in turn dependent upon the stage of differentiation reached. Nevertheless, growth is related to the duration of larval development (e.g. Wilbur & Collin, 1973) because conditions that are favorable for differentiation often are also favorable for growth (Smiths–Gill & Berven, 1979).

In conclusion, this study represents a first step towards determining the role of climatic factors in growth and development rates that are increased in response to the time constraint. One limitation of the study is that tadpoles can actively thermoregulate by moving between shallow and deep water, a fact that was not taken into consideration in this study. Nevertheless, the results suggest that temperatures below 20°C are too low to allow normal tadpole development and growth, while higher temperatures could favour growth and developmental conditions. In further studies it would be interesting to examine the existence of adaptive plasticity in development and growth rates in response to pond drying, as well as investigate whether a process of local adaptation is emerging in the study area.

#### Acknowledgements

We thank the Departament de Producció Vegetal i Ciència Forestal of University of Lleida, and especially Matilde Eizaguirre.

#### References

- Altwegg, R. & Reyer, H. U., 2003. Patterns of natural selection on size at metamorphosis in water frogs. *Evolution*, 57: 872–882.
- Álvarez, D. & Nicieza, G., 2002. Effects of temperature and food quality on anuran larval growth and metamorphosis. *Functional Ecology*, 16: 640–648.
- Atkinson, D., 1994. Temperature and organism size – a biological law for ectotherms? *Advances in Ecological Research*, 25: 1–58.
- 1996. Ectotherm life–history responses to developmental temperature. *Animals and Temperature. Phenotypic and Evolutionary Adaptation*: 183–204 (I. A. Johnston & A. F. Benett, Eds.). Cambridge Univ. Press.
- Berven, K. A. & Gill, D. E., 1983. Interpreting geographic variation in life history traits. *American Zoologist*, 23: 85–97.
- Brady, L. D. & Griffiths, R. A., 2000. Development responses to pond desiccation in tadpoles of the British anuran amphibians (*Bufo bufo*, *B. calamita* & *R. temporaria*). *Journal of Zoology*, 252: 61–69.
- Brattstrom, B. H., 1963. A preliminary review of thermal requirements of amphibians. *Ecology*, 44: 238–255.
- Brockelman, W. Y., 1969. An analysis of density effects and predation in *Bufo americanus* tadpoles. *Ecology*, 50: 632–644.
- Calef, G. W., 1973. Natural mortality of tadpoles in a population of *Rana aurora*. *Ecology*, 54: 741–758.
- Conesa, J. A., Mayoral, A., Pedro, J. & Recasens, J., 1994. *El paisatge vegetal dels espais d'interès naturals de Lleida: area meridional*. I.E.I. Diputació de Lleida.
- Etkin, W., 1964. Metamorphosis. In: *Physiology of the Amphibia*: 427–468 (J. A. Moore, Ed.). New York Academic Press, New York.
- Feder, M. E. & Burggren, W. W., 1992. *Environmental physiology of the amphibians*. Chicago, London, The Univ. of Chicago Press., Brockelman.
- Frost, D. R., Grant, T., Faivovich, J., Bain, R. H., Haas, A., Haddad, C. F. B., De Sá, R. O., Channing, A., Wilkinson, M., Donnellan, S. C., Raxworthy, C. J., Campbell, J. A., Blotto, B. L., Moler, P., Drewes, R. C., Nussbaum, R. A., Lynch, J. D., Green, D. M. & Wheeler, W. C., 2006. The amphibian tree of life. *Bulletin of American Museum Natural History*, 297: 1–370.
- Gosner, K. L., 1960. A simplified table for staging anuran embryos and larvae with notes on identification. *Herpetologica*, 16: 183–190.
- Hayes, T., Chan & Licht, P., 1993. Interactions of temperature and steroid on larval growth, development, and metamorphosis in a toad (*Bufo boreas*). *Journal of Experimental Zoology*, 299: 206–215.
- Laugen, A. T., Laurila, A. & Merilä, J., 2003a. Latitudinal and temperature dependent variation in embryonic development and growth in *Rana temporaria*. *Oecologia*, 135: 548–554.

- Laugen, A. T., Laurila, A., Räsänen, K. & Merilä, J., 2003b. Latitudinal countergradient variation in the common frog (*Rana temporaria*) development rates—evidence for local adaptation. *Journal of Evolutionary Biology*, 16: 996–1005.
- Loman, J., 1999. Early metamorphosis in common frog *Rana temporaria* tadpoles at risk of drying: an experimental demonstration. *Amphibia–Reptilia*, 20: 421–430.
- 2002. Temperature, genetic and hydroperiod effects on metamorphosis of brown frogs *Rana arvalis* and *R. temporaria* in the field. *Journal of Zoology*, 258: 115–129.
- Miaud, C., Sanuy, D. & Avrillier, J. N., 2000. Terrestrial movements of natterjack toad *Bufo calamita* (Amphibia, Anura) in semi-arid, agricultural landscape. *Amphibia–Reptilia*, 21: 357–144.
- Newman, R. A., 1987. Effects of density and predation on *Scaphiopus couchii* tadpoles in desert ponds. *Oecologia*, 71: 301–307.
- 1989. Developmental plasticity of *Scaphiopus couchii* tadpoles in an unpredictable environment. *Ecology*, 70: 1775–1787.
- Randall, D., Burggren, W. & French, K., 1997. *Eckert Animal Physiology: Mechanisms and Adaptations*. W. H. Freeman and Co., New York.
- Reques, R. & Tejedo, M., 1997. Reaction norms for metamorphic traits in natterjack toads to larval density and pond duration. *Journal of Evolutionary Biology*, 10: 829–851.
- Ryan, T. J. & Winne, C. T., 2001. Effects of hydroperiod on metamorphosis in *Rana sphenoccephala*. *American Midlands Naturalist*, 145: 46–53.
- Semlitsch, R. D., Scott, D. E. & Pechmann J. H. K., 1988. Time and size at metamorphosis related to adult fitness in *Ambystoma talpoideum*. *Ecology*, 69: 184–192.
- Smith, D. C., 1983. Factors controlling tadpole populations of the chorus frog (*Pseudacris triseriata*) on Isle Royale, Michigan. *Ecology*, 64: 501–510.
- Smith–Gill, S. J. & Berven, K. A., 1979. Predicting amphibian metamorphosis. *American Naturalist*, 113: 563–585.
- Tejedo, M. & Reques, R., 1994. Plasticity in metamorphic traits of natterjack tadpoles: the interactive effects of density and pond duration. *Oikos*, 71: 295–304.
- Thorntwaite, C. W. & Mather, J. R., 1955. The water balance. *Climatology*, 8: 1–104.
- Wilbur, H. M. & Collins, J. P., 1973. Ecological aspects of amphibian metamorphosis. *Science*, 182: 1305–1314.
-